Introduction to

# **Drug Disposition and Pharmacokinetics**









### Introduction to Drug Disposition and Pharmacokinetics

## Introduction to Drug Disposition and Pharmacokinetics

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### Preface

The application of knowledge of drug disposition, and skills in pharmacokinetics, is crucial to the development of new drugs and to a better understanding of how to achieve maximum benefit from existing ones. No matter how efficacious a potential new drug may be in the laboratory, the development of the new agent will come to nought if it cannot be delivered to its site of action and maintained there for sufficient duration for it to have the desired effect. Similarly, if it transpires that one (or more) of its metabolites is toxic then further development will cease. Also, knowledge of the concepts and principles of pharmacokinetics allows better clinical use of drugs and for this reason the topic is considered to some degree in most pharmacological texts.

We have written this book for those wishing, or needing, to study the topic in more depth than covered by most books of pharmacology. As an introductory text to drug disposition and pharmacokinetics, the book takes the reader from basic concepts to a point where those who wish to will be able to perform pharmacokinetic calculations and be ready to read more advanced texts and research papers. Exercises and further references have been included on the companion website for such readers.

The book will be of benefit to students of medicine, pharmacy, biomedical sciences and veterinary science, particularly those who have elected to study the topic in more detail, such as via electives and special study modules. It will be of benefit to those involved in drug discovery and product development, pharmaceutical and medicinal chemists, as well as budding toxicologists and forensic scientists who require the appropriate knowledge to interpret their findings. It will also provide a starting point for clinical pharmacologists, and may find use amongst professionals on the fringes of biomedical science, such as analytical chemists and patent lawyers. The book has been arranged to take the reader from a brief introduction to largely qualitative descriptions of drug disposition, and then to kinetic modelling to explain and predict drug effects, and to assist in the development of analytical skills that are needed in evaluating and mathematically describing pharmacokinetics. From there, we describe the special factors encountered in patient populations. The journey finishes with consideration of how aspects of drug interactions and toxicity are explainable, and indeed predictable, from consideration of the pharmacokinetic principles described in the previous chapters. The book concludes with a short history of the topic from its origins to where it is today and reflects upon the future, including the contribution that the reader might make. This chapter may be read as an introduction, but we feel the reader will gain most benefit by reading it with the knowledge of the previous chapters.

We envisage that some students will find the first chapters helpful in their early years and find the later ones more relevant if they gain more clinical experience, or find themselves working alongside clinicians. For some readers the book will provide all that they require but for others the book will be a valuable introduction to the topic, allowing them to progress to more advanced aspects of the subject.

We are grateful to the publishers for agreeing to the use of full-colour illustrations throughout as this enhances the learning experience and enables readers to assimilate what they need to know much more quickly. For example, many of the principal pharmacokinetic relationships can be demonstrated empirically by the movement of dye into and out of volumes of water, an approach we have used to illustrate the validity of several models, and one that has proved popular with our own students. We encourage the reader to visualize such movement, and we have reinforced this by including colour plates. Specific examples, some from our own experience, and others from the literature, often of seminal importance to the development of the subject, are included. Some of these examples refer to drugs that might be considered as being 'old', and indeed some may no longer be available in some countries, but many, propranolol, warfarin, digoxin, aspirin, and theophylline for instance, are still important members of their respective classes. Some of our examples, such as phenacetin and phenylbutazone, have been withdrawn or are no longer used in humans, but were the subject of investigations that informed our understanding, particularly of interactions and toxicity. We have not felt it necessary to search for examples involving more recently introduced drugs when drugs with more familiar names have already provided the examples that were needed. To do so would only increase the burden of drug names for the reader to learn.

In regard to drug names, some years ago the World Health Organization (WHO) started the introduction of a system of recommended international non-proprietary names (rINNs), which we have followed. Thus, different names and spellings may be encountered in different texts until the system is adopted universally. Generally it is clear as to what the new spelling refers, amfetamine and amphetamine, for instance, but when it is unclear we have added the alternative name in parentheses. This will be particularly helpful for our North American readers. Also, there is at present no internationally agreed system for pharmacokinetic symbols, but we have elected to use as simple a system as is possible, only adding additional sub- and superscripts when the meaning is not clear from the context.

Chapter 1 introduces the reader to the fundamental concepts related to the growth and decay of drug concentrations in plasma occurring alongside the growth and decay of effects, developed further in later chapters, and then continues with a brief presentation of the general chemical principles underlying the key mechanisms and processes described in the later chapters. To some extent, this reviews the relevant scientific language needed. Chapters 2 and 3 are largely qualitative descriptions of how drugs are administered and the physiochemical properties that influence their absorption, distribution, metabolism and excretion. Pharmacokinetic modelling of drugs and their metabolites can be found in Chapters 4-6. Chapter 7 examines extraction and clearance in more detail as these are the cornerstones of modern approaches to the subject and are vital to understanding physiologically-based pharmacokinetic modelling. Chapter 8 is devoted to the integration of pharmacokinetics and pharmacodynamics (PK-PD), while Chapter 9 describes the pharmacokinetics of what we have referred to as 'large molecules', a topic that is becoming increasingly important with the recent introduction of therapeutic enzymes and monoclonal antibodies. The next three chapters (10-12) deal with what can be referred to as 'special populations' or 'special considerations', sex, disease, age and genetics in particular. The plan in these chapters is that the reader will develop the ability to relate the many variables that affect drug response in the whole animal to the standard pharmacokinetic patterns shown in Figure 1.1 and described in Chapter 4, and later. Chapter 13 exemplifies the importance of pharmacokinetics in considering aspects of drug interactions and toxicity. Thus our sequence is from scientific preparation, through relevant science, to an introduction to clinical applications.

Additional material can be found on the companion website. This includes multiple choice questions, calculations and simulation exercises. We have also included suggestions for individual research or, preferably, for group discussions. Some of the material for these may be found in our previous book, *Drug Disposition and Pharmacokinetics: From Principles to Applications*. This also contains chapters on extrapolation from animals to humans and therapeutic drug monitoring, which we elected not to include in this book.

Whatever your reason for choosing this book we hope that reading it proves to be both a beneficial and an enjoyable experience.

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### **Companion Website Directions**

The companion website associated with this book (http://www.wiley.com/go/curryand whelpton/IDDP) contains a number of questions, exercises, simulations and more detailed explanations of selected topics which we feel readers will find helpful. We have taken this approach to encourage learning beyond that which is possible using the book on its own. Thus the website materials will assist you in improving your competence in the simple calculations that are essential to the subject, and also lead you to utilize other literature to broaden the educational experience. Hopefully, it will stimulate thinking, both by the reader on his or her own, and in the group discussions that we have found to be very helpful in the discovery of 'grey' areas of the subject, and which can facilitate understanding of controversial issues.

The resources are displayed in two ways:

- 1. By chapter: Use the drop-down menu at the top to view resources for that chapter.
- 2. By resource: Click the name of the resource in the top menu to see all content for that resource.

The types of resource include:

- *Multiple choice questions*: so that readers can quickly check their knowledge and comprehension; these can also be used as discussion topics.
- *Simulations*: so that the effects of changing pharmacokinetic parameters may be observed.
- Short questions: requiring brief explanations and sketches.
- *Topics for group discussion or private research:* these require students to think and research using more advanced texts and original scientific papers, and to reflect generally on what they have learnt.

Answers to the questions and, where appropriate, worked examples are provided, so that individuals using the book as a self-teaching aid can monitor their progress. Course leaders should be able to devise suitable questions for enhancing learning, and for testing their students.

We have suggested initial values for the simulations, which have been designed to reinforce the material in the book and should prove useful to all readers (see Appendix 4). Additionally, curve fitting using the method of residuals is demonstrated using a spreadsheet that can be used as a template for other sets of data. There is a detailed presentation explaining the relationship between apparent volumes of distribution in multiple-compartment models.

Should students require more examples, these can be accessed via the companion website that accompanies *Drug Distribution and Pharmacokinetics: From Principles to Applications*:

www.wiley.com/go/curryandwhelpton/IDDP

### 1

### **Introduction: Basic Concepts**

#### Learning objectives

This chapter was written for those unfamiliar with certain aspects of pharmacology and chemistry, including physical chemistry, and for those who feel a little revision would be helpful. By the end of the chapter the reader should be able to:

- use the Henderson–Hasselbalch equation to calculate the ionization of weak acids and bases
- plot concentration-time data to determine first-order and zero-order rate constants
- explain the effect of ionization on the partitioning of weak electrolytes between buffers and octanol.

#### **1.1 Introduction**

Pharmacology can be divided into two major areas, pharmacodynamics (PD) – the study of what a drug does to the body – and pharmacokinetics (PK) – the study of what the body does to the drug; hardly rigorous definitions but they suffice. Drug disposition is a collective term used to describe drug absorption, distribution, metabolism and excretion whilst pharmacokinetics is the study of the rates of these processes. By subjecting the observed changes, for example in plasma concentrations as a function of time, to mathematical

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#### 2 Introduction to Drug Disposition and Pharmacokinetics

equations (models) pharmacokinetic parameters such as elimination half-life  $(t_{y_2})$ , volume of distribution (*V*) and plasma clearance (*CL*) can be derived. Pharmacokinetic modelling is important for the:

- · selection of the right drug for pharmaceutical development
- evaluation of drug delivery systems
- design of drug dosage regimens
- appropriate choice and use of drugs in the clinic.

A detailed knowledge of mathematics is not required to understand pharmacokinetics and it is certainly not necessary to be able to differentiate or integrate complex equations. The few examples in this book are standard differentials or integrals that can be quickly learnt if they are not known already. To understand the equations in this book requires little more than a basic knowledge of algebra, laws of indices and logarithms, a brief explanation of which can be found in Appendix 1. Furthermore, the astute reader will quickly realize that, although seemingly different, many equations take the same form, making learning easier. For example, drug binding to macromolecules, whether they be receptors, plasma proteins, transporters or enzymes, can be described using the same basic equation. Similarly, the equation describing the time course of formation and excretion of a drug metabolite is very much like that describing the plasma concentrations during the absorption and elimination of a drug.

The role of pharmacokinetics is illustrated in Figure 1.1. There is an optimum range of concentrations over which a drug has beneficial effects, but little or no toxicity – this range is the therapeutic range, sometimes referred to as the therapeutic window. There is a threshold concentration below which the drug in ineffective and a higher threshold above which adverse effects become problematic. If a single dose of a drug, for example aspirin taken to relieve a headache, is consumed, the concentration in the plasma will rise until the aspirin becomes effective. After a period of time the processes which remove aspirin from the body will reduce the concentration until the drug is no longer effective (Figure 1.1, curve (a)).



**Figure 1.1** Typical concentration–time curves after oral administration of a drug: (a) single dose of drug; (b) a single dose twice the size of the previous one; (c) the same drug given as divided doses. The dose and frequency of dosing for (c) were calculated to ensure the concentrations remained in the therapeutic window.

The short duration of action may be fine for treating a headache but if the aspirin is to treat rheumatoid arthritis a much longer duration of action is required. Simply increasing the size of the dose is not the answer because eventually the plasma concentrations will enter the toxic region (Figure 1.1, curve (b)). However, by giving the aspirin as smaller divided doses at regular intervals the plasma concentrations can be maintained within the therapeutic window (Figure 1.1, curve (c)). The three curves depicted in Figure 1.1 were produced using relatively simple pharmacokinetic equations which will be explained later.

#### 1.2 Drugs and drug nomenclature

A drug is a substance that is taken, or administered, to produce an effect, usually a desirable one. These effects are assessed as physiological, biochemical or behavioural changes. There are two major groups of chemicals studied and used as drugs. First, there is a group of pharmacologically interesting endogenous substances, for example epinephrine, insulin and oxytocin. Second, there are the non-endogenous or 'foreign' chemicals (xenobiotics), which are mostly products of the laboratories of the pharmaceutical industry. Early medicines, some of which have been used for at least 5000 years, relied heavily on a variety of mixtures prepared from botanical and inorganic materials. Amongst the plant materials, the alkaloids, morphine from opium, cocaine from coca leaves and atropine from the deadly nightshade (belladonna) are still used today. Insulin, once obtained from pigs (porcine insulin), is more usually genetically engineered using a laboratory strain of *Escherichia coli* bacteria to produce human insulin. A few inorganic chemicals are used as drugs, including lithium carbonate ( $Li_2CO_3$ ) and sodium hydrogen carbonate (sodium bicarbonate, NaHCO<sub>3</sub>).

#### 1.2.1 Drug nomenclature

Wherever possible specific drug examples are given throughout this book, but unfortunately drug names can lead to confusion. Generally a drug will have at least three names: a full chemical name, a proprietary name, i.e. a trade name registered to a pharmaceutical company, and a non-proprietary name (INN) and/or an approved or adopted name. Names that may be encountered include the British Approved Name (BAN), the European Pharmacopoeia (EuP) name, the United States Adopted Name (USAN), the United States Pharmacopoeia (USP) name and the Japanese Approved Name (JAN). The World Health Organization (WHO) is introducing a system of recommended INNs and it is hoped that this will became the norm for naming drugs, replacing alternative systems (http://www. who.int/medicines/services/inn/innguidance/en/, accessed 17 February 2016). For example, lidocaine is classed as a rINN, USAN and JAN, replacing the name lignocaine that was once a BAN. Generally, the alternatives obviously refer to the same drug, e.g. ciclosporin, cyclosporin and cyclosporine. There are some notable exceptions, for example pethidine is known as meperidine in the US and paracetamol as acetaminophen. Even a simple molecule like paracetamol may have several chemical names but the number of proprietary names or products containing paracetamol is even greater, including Panadol, Calpol, Tylenol and Anadin Extra. It is therefore necessary to use an unequivocal approved name whenever possible, but alternative names and spellings are likely to be encountered, some examples of which are given in Table 1.1. Useful sites for checking, names, synonyms,

#### 4 Introduction to Drug Disposition and Pharmacokinetics

| rINN:BAN                | USAN:USP            | Alternative spellings*   |
|-------------------------|---------------------|--------------------------|
| Aciclovir               | Acyclovir           | Acyclovir                |
| Amfetamine              | Amphetamine         | Amphetamine              |
| Bendroflumethiazide     | Bendroflumethiazide | Bendrofluazide           |
| Benzylpenicillin        | Penicillin G        | Benzyl penicillin        |
| Cefalexin               | Cephalexin          | Cephalexin               |
| Ciclosporin             | Cyclosporine        | Cyclosporin              |
| Epinephrine             | Epinephrine         | Adrenaline               |
| Furosemide              | Furosemide          | Frusemide                |
| Glycerol                | Glycerin            |                          |
| Glyceryl trinitrate     | Nitroglycerin       |                          |
| Indometacin             | Indomethacin        | Indomethacin             |
| Isoprenaline            | Isoproterenol       |                          |
| Lidocaine               | Lidocaine           | Lignocaine               |
| Metamfetamine           | Methamphetamine     | Methamphetamine          |
| Norepinephrine          | Norepinephrine      | Noradrenaline            |
| Paracetamol             | Acetaminophen       |                          |
| Pethidine               | Meperidine          |                          |
| Phenoxymethylpenicillin | Penicillin V        | Phenoxymethyl penicillin |
| Rifampicin              | Rifampin            | , , , ,                  |
| Salbutamol              | Albuterol           |                          |
| Sulfadimidine           | Sulfamethazine      | Sulphadimidine           |

 Table 1.1
 Differences in rINN and USAN nomenclature

\* Chiefly previous BAN entries.

chemical properties and the like include http://chem.sis.nlm.nih.gov/chemidplus/and www.chemicalize.org/ (accessed 17 February 2016).

#### 1.3 Law of mass action

The reversible binding of drugs to macromolecules such as receptors and plasma proteins is described by the law of mass action: 'The rate at which a chemical reaction proceeds is proportional to the active masses (usually molar concentrations) of the reacting substances'. This concept is easily understood if the assumption is made that for the reaction to occur, collision between the reacting molecules must take place. It follows that the rate of reaction will be proportional to the molar concentrations of the reacting molecules. If a substance X is transformed into substance Y,

$$X \rightarrow Y$$

the rate of reaction = k[X], where k is the rate constant and [X] represents the molar concentration of X at that time. If two substances A and B are reacting to form two other substances C and D, and if the concentrations of the reactants at any particular moment are [A] and [B] then:

$$A + B \rightarrow C + D$$

and the rate of reaction = k[A][B].

#### **1.3.1** Reversible reactions and equilibrium constants

Consider the reaction:

$$A + B \rightleftharpoons C + D$$

The rate of the forward reaction is:

forward rate = 
$$k_1$$
 [A][B] (1.1)

whilst the backward rate is:

backward rate = 
$$k_{-1}$$
 [C][D] (1.2)

where  $k_1$  and  $k_{-1}$  are the rate constants of the forward and backward reactions, respectively. When equilibrium is reached the forward and backward rates are equal, so:

$$k_{1}[\mathbf{A}][\mathbf{B}] = k_{-1}[\mathbf{C}][\mathbf{D}]$$
(1.3)

The equilibrium constant *K* is the ratio of the forward and backward rate constants, and rearranging Equation 1.3 gives:

$$K = \frac{k_1}{k_{-1}} = \frac{\left[\mathbf{C}\right]\left[\mathbf{D}\right]}{\left[\mathbf{A}\right]\left[\mathbf{B}\right]}$$
(1.4)

The term *dissociation constant* is used when describing the equilibrium of a substance which dissociates into smaller units, as in the case, for example, of an acid (Section 1.4). The term is also applied to the binding of a drug, D, to a macromolecule such as a receptor, R, or plasma protein (Sections 2.5 and 8.2). The complex DR dissociates:

$$DR \rightleftharpoons D + R$$

so:

$$K = \frac{\left[\mathbf{D}\right]\left[\mathbf{R}\right]}{\left[\mathbf{D}\mathbf{R}\right]} \tag{1.5}$$

An association constant is the inverse of a dissociation constant.

#### **1.3.1.1** Sequential reactions

When a product D arises as a result of several sequential reactions (Figure 1.2), it cannot be formed any faster than the rate of at which its precursor C is formed, which in turn cannot be formed any faster than its precursor B. The rates of each of these steps are determined by the rate constants  $k_1$ ,  $k_2$  and  $k_3$ , therefore, the rate at which D is formed will be the rate of the slowest step, i.e. the reaction with the lowest value of rate constant. Say, for example,  $k_2$  is the lowest rate constant, then the rate of formation of D is determined by  $k_2$  and the reaction  $B \rightarrow C$  is said to be the *rate-limiting* or *rate-determining* step. This concept is fundamental to understanding sustained-release preparations (Chapter 4) and also drug metabolism when it occurs in more than one step (Chapter 6).



*Figure 1.2* Illustration of the rate-determining step in sequential reactions. The thinnest arrow represents the smallest rate constant and is therefore the rate-determining step.

#### 1.3.2 Reaction order and molecularity

The order of a reaction is the number, n, of concentration terms affecting the rate of the reaction, whereas molecularity is the number of molecules taking part in the reaction. The order of a reaction is measured experimentally and because it is often close to an integer, 0, 1, or 2, reactions may be referred to as zero-, first- or second-order, respectively. The reaction

$$X \rightarrow Y$$

is clearly monomolecular, and may be either zero- or first-order depending on whether the rate is proportional to $[X]^0$  or  $[X]^1$ . The reactions

 $2X \rightarrow Y$ 

and

$$A + B \rightarrow C + D$$

are both bimolecular and second-order providing the rate is proportional to  $[X]^2$  in the first case and to [A][B] in the second. Note how the total reaction order is the sum of the indices of each reactant: rate  $\propto [A]^1[B]^1$ , so n=2. However, if one of the reactants, say A, is present in such a large excess that there is no detectable change in its concentration, then the rate will be dependent only on the concentration of the other reactant, B. Thus, the rate is proportional to  $[A]^0[B]^1$ . The reaction is first-order (rate  $\propto [B]$ ) but it is still bimolecular. Hydrolysis of an ester in dilute aqueous solution is a commonly encountered example of a bimolecular reaction which is first-order with respect to the concentration of ester and zero-order with respect to the concentration of water, giving an overall reaction order of unity.

Enzyme-catalysed reactions have reaction orders between 1 and 0 with respect to the drug concentration. This is because the Michaelis–Menten equation (Section 4.7) limits to zero-order when the substrate is in excess and the enzyme is saturated so that increasing the drug concentration will have no further effect on the reaction rate. When the concentration of enzyme is in vast excess compared to the substrate concentration, the enzyme concentration is not rate determining and the reaction is first order. Thus, the reaction order of an enzyme-catalysed reaction changes as the reaction proceeds and substrate is consumed.

#### 1.3.3 Decay curves and half-lives

As discussed above, the rate of a chemical reaction is determined by the concentrations of the reactants and from the foregoing it is clear that a general equation relating the rate of decline in concentration (-dC/dt), rate constant ( $\lambda$ ) and concentration (C) can be written:

$$-\frac{\mathrm{d}C}{\mathrm{d}t} = \lambda C^n \tag{1.6}$$

Note the use of  $\lambda$  to denote the rate constant when it refers to decay; the symbol is used for radioactive decay, when it is known as the decay constant. Use of  $\lambda$  to denote *elimina-tion* rate constants is becoming more prevalent in pharmacokinetic publications.

#### 1.3.3.1 First-order decay

Because first-order kinetics are of prime importance in pharmacokinetics, we shall deal with these first. For a first-order reaction, n=1 and

$$-\frac{\mathrm{d}C}{\mathrm{d}t} = \lambda C \tag{1.7}$$

Thus, the rate of the reaction is directly proportional to the concentration of substance present. As the reaction proceeds and the concentration of the substance falls, the rate of the reaction decreases. This is exponential decay, analogous to radioactive decay, where the probability of a disintegration is proportional to the number of unstable nuclei present. The first-order rate constant has units of reciprocal time (e.g.  $h^{-1}$ ). Integrating Equation 1.7 gives:

$$C = C_0 \exp(-\lambda t) \tag{1.8}$$

which is the equation of a curve that asymptotes to 0 from the initial concentration,  $C_0$  (Figure 1.3(a)). Taking natural logarithms of Equation 1.8 gives:

$$\ln C = \ln C_0 - \lambda t \tag{1.9}$$

which is the equation of a straight line of slope,  $-\lambda$  (Figure 1.3(b)). Before the advent of inexpensive calculators and the availability of spreadsheets, common logarithms were often used to plot log *C* against *t* when the slope was  $-\lambda/2.303$ . Another way of presenting the data is to plot *C* on a logarithmic scale. This approach was often used when computers where not readily available. The half-life can be read easily from such graphs and  $\lambda$  can be calculated via Equation 1.10.

The half-life  $(t_{y_2})$  is the time for the initial concentration  $(C_0)$  to fall to  $C_0/2$ , and substitution in Equation 1.9 gives:

$$t_{\frac{1}{2}} = \frac{\ln 2}{\lambda} = \frac{0.693}{\lambda}$$
(1.10)

because ln 2=0.693. This important relationship, where  $t_{y_2}$  is constant (independent of the initial concentration) and inversely proportional to  $\lambda$ , is *unique* to first-order reactions.



Figure 1.3 Curves for first-order decay plotted as (a) C versus t and (b) In C versus t.

Because  $t_{1/2}$  is constant, 50% is eliminated in  $1 \times t_{1/2}$ , 75% in  $2 \times t_{1/2}$  and so on. Thus, when five half-lives have elapsed less than 5% of the substance remains, and after seven half-lives less than 1% remains.

#### 1.3.3.2 Zero-order decay

For a zero-order reaction, n=0 and:

$$-\frac{\mathrm{d}C}{\mathrm{d}t} = \lambda C^0 = \lambda \tag{1.11}$$

Because  $C^0 = 1$ , it is clear that a zero-order reaction proceeds at a *constant rate*, and the zero-order rate constant must have units of *rate* (e.g. g L<sup>-1</sup> h<sup>-1</sup>). Integrating Equation 1.11:

$$C = C_0 - \lambda t \tag{1.12}$$

gives the equation of a straight line of slope,  $-\lambda$ , when concentration is plotted against time (Figure 1.4(a)). The ln *C* plot is a convex curve because initially the *proportion* of drug eliminated is less when the concentration is higher (Figure 1.4(b)).

The half-life can be obtained as before, and substituting  $t = t_{1/2}$  and  $C = C_0$  gives:

$$t_{y_2} = \frac{C_0}{2\lambda} \tag{1.13}$$

The zero-order half-life is inversely proportional to  $\lambda$ , as would be expected, but  $t_{\nu_2}$  is also directly proportional to the initial concentration. In other words, the greater the amount of drug present initially, the longer the time taken to reduce the amount present by 50%, as would be expected. The term 'concentration-dependent half-life' has been applied to this situation.

Equations such as Equations 1.8 and 1.12 are referred to as *linear* equations. Note that in this context it is important not to confuse 'linear' with 'straight-line'. While it is true



**Figure 1.4** Curves for zero-order decay plotted as (a) C versus t and (b)  $\ln C$  versus t. Note how for an initial concentration of 100,  $t_{10} = 25$ , but for an initial concentration of 50,  $t_{10} = 12.5$ .

that the equation of a straight-line is a linear equation, exponential equations are also linear. On the other hand, non-linear equations are those where the variable to be solved cannot be written as a linear combination of independent variables. The Michaelis–Menten equation is such an example.

#### 1.3.3.3 Importance of half-life in pharmacokinetics

Half-life is a very useful parameter in pharmacokinetics. It is much easier to compare the duration of action of drugs in terms of their relative half-lives rather than rate constants, and the rate of attainment of steady-state concentrations during multiple dosing and the fluctuations in peak and trough levels is a function of  $t_{1/2}$  (Figure 4.16). However, it is important to recognize at the outset that the half-life of a drug is, in fact, dependent on two other pharmacokinetic parameters, apparent volume of distribution (*V*) and clearance (*CL*). The apparent volume of distribution, as its name implies, is a quantitative measure of the extent to which a drug is distributed in the body (Section 2.4.1.1) whist clearance can be thought of as an indicator of how efficiently the body's eliminating organs remove the drug, therefore the larger the value of *CL*, the shorter will be  $t_{1/2}$ , and any change in half-life will be as a result of changes in either *V* or *CL*, or both (Section 4.2.1).

#### 1.4 Ionization

The degree of ionization of a molecule can have a major influence on its disposition and pharmacokinetics. The term *strength* when applied to an acid or base refers to its tendency to ionize. The term should not be confused with *concentrated*. Strong acids and bases can be considered to be 100% ionized at any practical pH value. Weak electrolytes, such as amines and carboxylic acids, are only partially ionized in aqueous solutions:

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$$R-NH_2 + H^{\textcircled{o}} \implies R-NH_3^{\textcircled{o}}$$
$$R-COOH \implies R-COO^{\textcircled{o}} + H^{\textcircled{o}}$$

The degree of ionization is determined by the  $pK_a$  of the ionizing group and the pH of the aqueous environment. The  $pK_a$  of the compound, a measure of its inherent acidity or basicity, is numerically equal to the pH at which the compound is 50% ionized.

For an acid, AH, dissolved in water:

$$AH \rightleftharpoons H^+ + A^-$$

The acid dissociation constant is:

$$K_{\rm a} = \frac{\left[\mathrm{H}^+\right]\left[\mathrm{A}^-\right]}{\left[\mathrm{A}\mathrm{H}\right]} \tag{1.14}$$

Clearly the more the equilibrium is to the right, the greater is the hydrogen ion concentration, with a subsequent reduction in the concentration of unionized acid, so the larger will be the value of  $K_a$ . Note that because  $pK_a$  is the negative logarithm of  $K_a$  (analogous to pH), strong acids have *low* values of  $pK_a$ . Taking logarithms (see Appendix 1 for details) of Equation 1.14 gives:

$$\log K_{a} = \log \left[ H^{+} \right] + \log \left[ A^{-} \right] - \log \left[ AH \right]$$
(1.15)

and on rearrangement:

$$-\log\left[\mathrm{H}^{+}\right] = -\log K_{\mathrm{a}} + \log \frac{\left[\mathrm{A}^{-}\right]}{\left[\mathrm{A}\mathrm{H}\right]}$$
(1.16)

Because  $-\log[H^+]$  is the pH of the solution:

$$pH = pK_{a} + \log \left[\frac{A^{-}}{[AH]}\right] = pK_{a} + \log \left[\frac{base}{[acid]}\right]$$
(1.17)

where  $pK_a = -\log K_a$ , by analogy with pH. Note that when  $[A^-] = [AH]$  the ratio is 1 and because  $\log(1)=0$ , the  $pK_a = pH$ , as stated earlier.

The range of  $pK_a$  values extends below 1 and above 14, but for the majority of drugs values are between 2 and 13. Benzylpenicillin ( $pK_a = 2.3$ ) is an example of a relatively strong acid and metformin ( $pK_a = 12.4$ ) is an example of a relatively strong base (Figure 1.5). It should be noted that it is not possible from a knowledge of the  $pK_a$  alone to say whether a substance is an acid or a base. It is necessary to know how the molecule ionizes. Pentobarbital,  $pK_a = 8.0$ , forms sodium salts and so must be an acid, albeit a rather weak one. The electron-withdrawing oxygen atoms result in the hydrogen atom being acidic. Diazepam,  $pK_a = 3.3$ , must be a base because it can be extracted from organic solvents into hydrochloric acid. Imines are weak bases because of delocalization of the nitrogen lone pair of electrons around the C=N double bond. In oxazepam, the

electron-withdrawing effect of the oxygen in the hydroxyl group reduces the  $pK_a$  (=1.7) of the imine compared to that in diazepam. Molecules can have more than one ionizable group, for example salicylic acid has a carboxylic acid ( $pK_a$ =3.0) and a weaker acidic phenol group ( $pK_a$ =13.4). The amide in oxazepam is very weakly acidic ( $pK_a$ =11.6), making this compound amphoteric, that is, both acidic and basic. Sulfonamides are usually more acidic than amides and if the primary aromatic amine is not acetylated they are amphoteric, as illustrated by sulfadimidine. Similarly, morphine is amphoteric, having a tertiary amine group ( $pK_a$ =8.0) and an acidic phenol ( $pK_a$ =9.9) (Figure 1.5).

Not all drugs ionize. The volatile and gaseous anaesthetics are usually neutral compounds, for example enflurane (CHCIF<sub>2</sub>-CF<sub>2</sub>C-O-CF<sub>2</sub>H), which is an ether. Alcohols such as ethanol and chloral hydrate (CCl<sub>3</sub>CH(OH)<sub>2</sub>) are usually referred to as being neutral as they do not ionize at physiological pH values.



**Figure 1.5** Examples of ionizable groups in selected drug examples. Note how the  $pK_a$  values range (strong to weak) from 2.3 to 11.6 for the acids and 12.4 to 1.7 for the basic groups. Acidic hydrogen atoms are shown in red and basic nitrogen atoms in blue.

#### 1.4.1 Henderson–Hasselbalch equation

Equation 1.17 is a form of the Henderson–Hasselbalch equation, which is important in determining the degree of ionization of weak electrolytes and calculating the pH of buffer solutions. If the degree of ionization is  $\alpha$ , then the degree non-ionized is  $(1 - \alpha)$  and, for an acid:

$$pH = pK_a + \log\frac{\alpha}{1 - \alpha}$$
(1.18)

Taking antilogarithms and rearranging allows the degree of ionization to be calculated:

$$\alpha = \frac{10^{(\text{pH}-\text{p}K_a)}}{1+10^{(\text{pH}-\text{p}K_a)}}$$
(1.19)

The equivalent equation for a base is:

$$\alpha = \frac{10^{(pK_a - pH)}}{1 + 10^{(pK_a - pH)}}$$
(1.20)

Although Equations 1.19 and 1.20 may look complex, they are easy to use. Using the ionization of aspirin as an example, the  $pK_a$  of aspirin is ~ 3.4, so at the pH of plasma (pH 7.4)

$$pH - pK_a = 7.4 - 3.4 = 4$$
$$\alpha = \frac{10^4}{1 + 10^4} = \frac{10000}{10001} = 0.9999$$

In other words aspirin is 99.99% ionized at the pH of plasma, or the ratio of ionized to non-ionized is 10,000:1. In gastric contents, pH 1.4, aspirin will be largely non-ionized; 1.4 - 3.4 = -2, so the ratio of ionized to non-ionized is  $1:10^{-2}$ , that is, there are 100 non-ionized molecules for every ionized one.

#### **1.5 Partition coefficients**

The ability of a drug to dissolve in, and so cross, lipid cell membranes can be a major factor in its disposition. This ability can be assessed from its partition coefficient. When an aqueous solution of a substance is shaken with an immiscible solvent (e.g. diethyl ether) the substance is extracted into the solvent until equilibrium between the concentration in the organic phase and the aqueous phase is established. For dilute solutions the ratio of concentrations is known as the distribution, or partition coefficient, P:

$$P = \frac{\text{concentration in organic phase}}{\text{concentration in aqueous phase}}$$
(1.21)

Organic molecules with large numbers of paraffin chains, aromatic rings and halogens tend to have large values of P, whilst the introduction of polar groups such as hydroxyl or carbonyl groups generally reduces the partition coefficient. Drugs with high partition

coefficients are lipophilic or hydrophobic, whereas those that are very water soluble and are poorly extracted by organic solvents are hydrophilic. Lipophilicity can have a major influence on how a drug is distributed in the body, its tendency to bind to macromolecules such as proteins and, as a consequence, drug activity. A relationship between partition coefficient and pharmacological activity was demonstrated as early as 1901, but it was in the 1960s that Corwin Hansch used regression analysis to correlate biological activity with partition coefficient. He chose *n*-octanol as the organic phase and this has become the standard for such studies (Figure 1.6). Because *P* can vary between <1 (poorly extracted by the organic phase) to several hundred thousand, values are usually converted to log *P*, to encompass the large range (Appendix 1).

#### 1.5.1 Effect of ionization on partitioning

Generally, ionized molecules cannot be extracted into organic solvents, or at least not appreciably. Thus, for weak electrolytes the amount extracted will be dependent on the degree of ionization, which of course is a function of the pH of the aqueous solution and the  $pK_a$  of the ionizing group, as discussed above (Section 1.4), and the partition coefficient. If the total concentration (ionized+non-ionized) of solute in the aqueous phase is measured and used to calculate an apparent partition coefficient D, then the partition coefficient P, can be calculated. For an acid:



$$P = D\left[1 + 10^{\left(pH - pK_{a}\right)}\right]$$
(1.22)

**Figure 1.6** (a) Partitioning of chloral hydrate is unaffected by buffer pH. (b) Partitioning of non-ionized amfetamine remains constant, 63:1. However, the ratio of ionized to non-ionized is affected by buffer pH and as a consequence affects the apparent partition coefficient (APC) and the proportion extracted (inset). Note how when  $pH=pK_{a'} x=1$  and there are equal concentrations of ionized and non-ionized amfetamine in the aqueous phase.

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and for a base:

$$P = D\left[1 + 10^{(\mathsf{pK}_a - \mathsf{pH})}\right] \tag{1.23}$$

When  $pH = pK_a$  then, because  $10^0 = 1$ , P = 2D. When the pH is very much less than the  $pK_a$ , in the case of acids, or very much larger than the  $pK_a$ , in the case of bases, there will be no appreciable ionization and then D will be a good estimate of P (Figure 1.6(b)).

Unless stated otherwise, log *P* is taken to represent the logarithm of the true partition coefficient, that is, when there is no ionization of the drug. However, for some weak electrolytes, biological activity may correlate better with the partition coefficient between octanol and pH7.4 buffer solution. These values are referred to as log *D* or log  $D_{74}$ .

Differences in the pH of different physiological environments, for example plasma and gastric contents, can have a major influence on the way drugs are absorbed and distributed (Chapter 2).

#### Summary

This chapter has introduced rates, rate constants and half-lives, all crucially important to understanding the pharmacokinetics described in Chapters 4 and 5, and elsewhere in this book. Rate-determining reactions will be encountered when considering sustained-release formulations (Chapter 4) and the kinetics of metabolism (Chapter 6). The influence of the degree of ionization on partitioning of weak electrolytes will be helpful in understanding Chapters 2 and 3.

#### **1.6 Further reading**

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### 2

### Drug Administration and Distribution

#### Learning objectives

By the end of this chapter the reader should be able to:

- describe the mechanisms by which substances cross biological membranes
- explain the pH-partition hypothesis and its role in the absorption and sequestration of weak acids and bases
- compare the advantages and disadvantages of administering drugs by the routes described in this chapter
- discuss apparent volume of distribution and explain its relationship with known anatomical volumes
- explain the mechanisms by which drugs are sequestered in tissues
- discuss plasma protein binding and explain its importance in pharmacology.

#### 2.1 Introduction

In order to achieve its effect, a drug must first be presented in a suitable form at an appropriate site of *administration*. It must then be *absorbed* from the site of administration and *distributed* through the body to its site of action. For the effect to wear off the drug must nearly always be *metabolized* and/or *excreted*. These processes are often given

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the acronym ADME, or occasionally LADME, where L stands for liberation of drug from its dosage form, which can be an important factor in the usefulness of the drug. Finally, drug residues are *removed* from the body. Removal refers to loss of material, unchanged drug and/or metabolic products, in urine and/or faeces, once this material has been excreted into the bladder or bowel by the kidneys and liver. Absorption and distribution comprise the *disposition* (placement around the body) of a compound. Metabolism and excretion comprise the *fate* of a compound.

The most common pathway for an orally administered drug is as indicated by the red arrows in Figure 2.1. This pathway involves metabolism and excretion of both unchanged drug and metabolites. A drug that is excreted in its unmetabolized form will bypass metabolism (orange). An intravenously administered drug undergoes no absorption (purple) but will be distributed, metabolized and excreted in the same way as it would if it had been orally administered. An oral dose may not be absorbed into the systemic circulation because it cannot cross the gastric mucosa (blue) or because it is rapidly converted to its metabolites in the intestinal mucosa or the portal circulation so that only the drug metabolites reach the systemic circulation; this is referred to as 'pre-systemic' or first-pass metabolism (Section 2.4.1.1). Excretion products in the intestine may be reabsorbed (green), for example as with enterohepatic cycling (Section 3.3.8.1), rather than being removed via the faeces.



*Figure 2.1* General scheme showing the relationship between the various events involved in drug disposition and fate (see text for details).

#### 2.2 Drug transfer across biological membranes

Absorption, distribution and excretion of drugs involve transfer of drug molecules across various membranes, such as the gastrointestinal (GI) epithelium, the renal tubular epithelium, the blood-brain barrier and the placental membrane. Transfer of

substances across biological membranes can occur by one or more of five possible mechanisms:

- Passive diffusion: through the membrane, down a concentration gradient.
- Filtration: through channel proteins and pores in the membrane.
- *Facilitated diffusion:* a carrier-mediated process that does not require energy and where the net flow is down a concentration gradient.
- *Active transport:* involving carrier processes, requiring energy and occurring against the concentration gradient.
- *Pinocytosis:* microscopic invaginations of the cell wall engulf drops of extracellular fluid and solutes are carried through in the resulting vacuoles of water.

Historically, passive diffusion has been viewed as by far the most important mechanism for foreign molecules (Section 2.2.1). Filtration can be transcellular, via pores in the plasma membrane (Figure 2.2) or paracellular (i.e. between cells) and is important for the transfer of small molecules into interstitial fluid via the fenestrations in peripheral capillaries (Section 2.2.3). Filtration plays a major part in the urinary excretion of drug molecules.



*Figure 2.2* Schematic representation of some of the mechanisms by which substances are transferred across lipid membranes.

There has been increased interest in the roles of active transport and facilitated diffusion in the disposition of drugs (Section 2.2.2) and the carrier proteins are being investigated as targets for new drugs. The transfer of small foreign compounds across membranes by pinocytosis is largely unknown, but it is believed to be the mechanism by which macromolecules such as botulinum toxin are absorbed. Pinocytosis and other forms of endocytosis are receiving a great deal of interest as a way of absorbing drugs as nanoparticles, micelles and the like. The ileum differs from the duodenum and the jejunum in that it has regions of aggregated lymphoid nodules known as Peyer's patches, and macromolecules, including antigens, can be absorbed via associated specialized cells, known as microfold, or M, cells, which one day may prove to be a route by which insulin can be administered orally (Lopes *et al.*, 2014). Anything that is absorbed in this way drains into the lymphatic system.

#### 2.2.1 Passive diffusion

The rate of diffusion of a drug, dQ/dt, is a function of the concentration gradient across the membrane,  $\Delta C$ , the surface area over which transfer occurs, A, the thickness of the membrane,  $\Delta x$ , and the diffusion coefficient, D, which is characteristic of the particular drug. The diffusion constant of a compound is a function of its solubility in the membrane, its molecular weight and its steric configuration, therefore:

$$\frac{\mathrm{d}Q}{\mathrm{d}t} = RDA\frac{\Delta C}{\Delta x} \tag{2.1}$$

where R is the partition coefficient of the drug between the membrane and the aqueous phase. Because for a given drug diffusing through a particular membrane A, D, R and the distance over which diffusion is occurring are constants, these can be combined into one constant, known as permeability, P, so that:

$$\frac{\mathrm{d}Q}{\mathrm{d}t} = P\Delta C = P\left(C_1 - C_2\right) \tag{2.2}$$

It is important to note that not only is the rate of diffusion dependent of the nature of the drug and the membrane and, of course, the concentration gradient, the area over which transfer is taking place is of major importance. The micro villi of the small intestine and the kidney proximal convoluted tubular cells provide a large surface area over which transfer can take place.

#### 2.2.1.1 pH-partition hypothesis

With passive diffusion drug molecules have to dissolve in the membrane and so low molecular weight, lipophilic species diffuse freely, but polar, particularly ionized, molecules do not. Whether a weak electrolyte is ionized or not will be a major determinant of whether it will diffuse across a biological membrane. The ratio of ionized to non-ionized forms is a function of the pH of the aqueous environment and the acid dissociation constant,  $K_{a,}$  as explained in Section 1.6. This means that in any aqueous solution both ionized and non-ionized forms are present to varying degrees.

Calculation of the equilibrium distribution of aspirin between stomach contents and plasma water illustrates the importance of pH partitioning (Figure 2.3). It will be readily appreciated that, at equilibrium, for the *non-ionized* species (which alone passes the lipid membrane) the concentration ratios will be the same:

| concentration in membrane     | concentration in membrane     |
|-------------------------------|-------------------------------|
| concentration in gastric acid | concentration in plasma water |



**Figure 2.3** Equilibrium distribution of aspirin ( $pK_a = 3.4$ ) between two solutions of pH 1.4 and pH 7.4 separated by a simple membrane. Relative concentrations are shown in [].

Thus for the non-ionized species the concentrations on each side of the membrane will be identical. For convenience, in Figure 2.3 we have designated this concentration as being 100 units, [100]. However, from the Henderson–Hasselbalch equation (Section 1.6.1) the ratio of ionized to non-ionized aspirin at pH 1.4 is 1:100. In other words, for every 100 units non-ionized there is 1 unit of ionized aspirin. In the alkaline environment of the plasma, aspirin will be highly ionized, the ratio being 10,000:1, so at pH 7.4, for every 100 molecules of non-ionized aspirin there will be 10,000 × 100 = 1,000,000 molecules ionized, therefore at equilibrium the ratio of total material on the two sides will be:

$$\frac{\text{total concentration in acid}}{\text{total concentration in plasma}} = \frac{100 + 1}{1,000,000 + 100} = \frac{101}{1,000,100} \approx \frac{1}{9900}$$

that is, almost 10,000 to 1 in favour of plasma. If drug molecules are introduced into any part of the system, they will transfer between the various media, including the membrane, until this concentration ratio is achieved. However, when aspirin is ingested, after absorption into the plasma it will be carried away in the bloodstream. Any binding to plasma proteins will also increase the concentration gradient in favour of absorption because there will be fewer non-ionized, non-bound molecules in the plasma.

#### 2.2.2 Carrier-mediated transport

There are two important superfamilies of drug transporters, solute carriers (SLC) and ABC transporters, the latter functioning chiefly as efflux pumps (Table 2.1).

The SLC superfamily of solute carriers is the second largest family of membrane proteins after G protein-coupled receptors. The family includes the organic cation transporters

| Transporter        | Gene    | Selected localizations                                                                                                            | Selected substrates                                              | Selected inhibitors                                                                  |
|--------------------|---------|-----------------------------------------------------------------------------------------------------------------------------------|------------------------------------------------------------------|--------------------------------------------------------------------------------------|
| OATP1A2            | SLCO1A2 | Brain<br>Enterocytes (luminal)<br>Kidnev PCT (luminal)                                                                            | Atenolol<br>Ciprofloxacin<br>Fexofenadine                        | Grapefruit juice<br>Orange juice<br>Verapamil                                        |
| OATP2B1            | SLCO2B1 | Enterocytes (luminal)<br>Hepatocytes<br>(sinusoidal)                                                                              | Aliskiren<br>Atorvastatin<br>Pravastatin                         | Ciclosporin<br>Grapefruit juice<br>Orange juice                                      |
| OATP1B1            | SLCO1B1 | Hepatocytes<br>(sinusoidal)                                                                                                       | Pravastatin<br>Rifampicin<br>Simvastatin                         | Ciclosporin<br>Lopinavir<br>Rifampicin                                               |
| OATP1B3<br>(OATP8) | SLCO1B3 | Hepatocytes<br>(sinusoidal)<br>Kidney PCT<br>(basolateral)                                                                        | Paclitaxel<br>Pravastatin<br>Rifampicin                          | Clarithromycin<br>Ciclosporin<br>Rifampicin                                          |
| OAT1               | SLC22A6 | (basolateral)<br>(basolateral)                                                                                                    | Captopril<br>Cidofovir<br>Furosemide                             | Indometacin<br>Probenecid                                                            |
| OAT3               | SLC22A8 | Kidney PCT<br>(basolateral)                                                                                                       | Benzylpenicillin<br>Furosemide<br>Methotrexate<br>Pravastatin    | Indometacin<br>Probenecid<br>Salicylate                                              |
| OCT2               | SLC22A2 | Brain<br>Lung<br>Kidney PCT<br>(basolateral)                                                                                      | Cimetidine<br>Cisplatin<br>Metformin<br>Ranitidine               | Cimetidine<br>Procainamide<br>Ranitidine<br>Trimethoprim                             |
| MATE1              | SLC47A1 | Hepatocytes<br>(canalicular)<br>Kidney PCT<br>(basolateral)<br>Skeletal muscle                                                    | Cefalexin<br>Metformin<br>Procainamide                           | Cimetidine<br>Pyrimethamine<br>Quinidine                                             |
| PepT1              | SLC15A1 | Enterocytes (luminal)<br>Kidney PCT (luminal)                                                                                     | Penicillins<br>Cephalosporins<br>Enalapril<br>Valaciclovir       | Glibenclamide<br>Tolbutamide                                                         |
| P-gp               | ABCB1   | Adrenal gland<br>Blood-brain barrier<br>Enterocytes (luminal)<br>Hepatocytes<br>(canalicular)<br>Placenta<br>Kidnov PCT (luminal) | Digoxin<br>Docetaxel<br>Fexofenadine<br>Ritonavir<br>Vinblastine | Clarithromycin<br>Ciclosporin<br>Itraconazole<br>Quinidine<br>Ritonavir<br>Verapamil |
| MRP2               | ABCC2   | Enterocytes (luminal)<br>Hepatocytes<br>(canalicular)<br>Kidney PCT (luminal)                                                     | Glucuronide and<br>GSH conjugates<br>Bromosulfophthalein         | Ciclosporin<br>Probenecid                                                            |
| BCRP               | ABCG2   | Enterocytes (luminal)<br>Hepatocytes<br>(canalicular)<br>Mammary glands                                                           | Doxorubicin<br>Methotrexate<br>5-Flurouracil                     | Ritonavir<br>Omeprazole                                                              |

 Table 2.1
 Selected examples of SLC and ABC transporters

BCRP: breast cancer resistance protein; MATE: multidrug and toxin extrusion protein; MRP: multidrug resistanceassociated protein; OAT: organic anion transporter; OATP: organic anion-transporting polypeptide; OCT: organic cation transporter; PepT: peptide transporter; P-gp: P-glycoprotein.