

Farida Nissan-Azzouz

**Fine Mapping of the Barley Locus *Rym11*
Conferring Resistance to the Barley Yellow
Mosaic Virus Complex**



Cuvillier Verlag Göttingen

Aus dem Institut für Pflanzenbau & Pflanzenzüchtung I
der Justus-Liebig-Universität Giessen
Lehrstuhl für Pflanzenzüchtung
Leiter: Prof. Dr. Dr.h.c. Wolfgang Friedt

**Fine Mapping of the Barley Locus *Rym11*
Conferring Resistance to the
Barley Yellow Mosaic Virus Complex**

Dissertation

zur Erlangung des Doktorgrades (Dr. agr.) beim
Fachbereich Agrarwissenschaften, Ökotoxikologie und Umweltmanagement
der Justus-Liebig-Universität Giessen

vorgelegt von
Farida Nissan-Azzouz

Giessen 2004

Bibliografische Information Der Deutschen Bibliothek

Die Deutsche Bibliothek verzeichnet diese Publikation in der Deutschen Nationalbibliografie; detaillierte bibliografische Daten sind im Internet über <http://dnb.ddb.de> abrufbar.

1. Aufl. - Göttingen : Cuvillier, 2004

Zugl.: Giessen, Univ., Diss., 2004

ISBN 3-86537-071-3

Vorsitzender: Prof. Dr. Dr. h.c. W. Opitz von Boberfeld

Gutachter: Prof. Dr. Dr. h.c. W. Friedt

Gutachter: Prof. Dr. K.-H. Kogel

Prüfer: Prof. Dr. B. Honermeier

Prüfer: Prof. Dr. S. Schnell

Tag der mündlichen Prüfung: 9. Februar 2004

© CUVILLIER VERLAG, Göttingen 2004

Nonnenstieg 8, 37075 Göttingen

Telefon: 0551-54724-0

Telefax: 0551-54724-21

www.cuvillier.de

Alle Rechte vorbehalten. Ohne ausdrückliche Genehmigung des Verlages ist es nicht gestattet, das Buch oder Teile daraus auf fotomechanischem Weg (Fotokopie, Mikrokopie) zu vervielfältigen.

1. Auflage, 2004

Gedruckt auf säurefreiem Papier

ISBN 3-86537-071-3

For my Ramsey

1 Introduction	1
1.1 Disease resistance in plants	1
1.1.1 Broad-spectrum resistance	1
1.1.2 Hypersensitivity resistance	2
1.1.3 Quantitative resistance	3
1.2 Fine mapping: a prerequisite for resistance gene cloning	4
1.3 Molecular markers	7
1.4 The barley yellow mosaic virus complex	9
1.4.1 The disease, its relevance and causal agents	9
1.4.2 Genetics of resistance	11
1.5 Barley: importance, genomics and genetic mapping	13
2 Objectives	18
3 Material and Methods	17
3.1 Plant material	17
3.1.1 'Marinka' x 'PI1963' population	17
3.1.2 Mapping population	17
3.1.3 Disease assessment	17
3.1.4 DNA extraction	18
3.1.5 DNA concentration measurement	19
3.2 Molecular analysis	20
3.2.1 Development of random amplified polymorphic DNA (RAPD) markers	20
3.2.1.1 RAPD amplification	21
3.2.1.2 Polymorphism detection	22
3.2.2 Mapping of simple sequence repeats (SSR) markers	23
3.2.2.1 SSR amplification	25
3.2.2.2 Polymorphism detection	26

3.2.3 Saturation with amplifical fragment length polymorphism (AFLP) markers	28
3.2.3.1 Bulked segregant analysis	28
3.2.3.2 AFLP technology	28
3.2.3.3 Polymorphism detection	32
3.3 Data analysis	33
3.3.1 Marker segregation test	33
3.3.2 Linkage analysis	34
4 Results	36
4.1 RAPD development	36
4.1.1 Polymorphism screening	36
4.1.2 RAPD mapping	38
4.1.3 RAPD marker transfer to <i>rym11</i> mapping populations	39
4.1.3.1 Polymorphism level	39
4.1.3.2 Linkage analysis	39
4.2 SSR mapping	42
4.2.1 Polymorphism level	42
4.2.2 Resistance gene <i>rym11</i> and marker segregation	44
4.2.3 Linkage analysis	44
4.3 AFLP saturation	48
4.3.1 AFLP informativeness	48
4.3.2 Bulk design	48
4.3.3 Identification of linked AFLPs	49
4.3.4 Linkage analysis	52
5 Discussion	56
5.1 Resistance gene <i>rym11</i> and marker segregation	56
5.2 Resistance gene <i>rym11</i> : chromosomal location	57
5.3 SSR map accuracy	58

5.4 Bulk design for RAPD development and AFLP saturation	61
5.5 Co-segregating AFLP loci as potential markers for positional cloning	63
5.6 Tightly-linked SSRs and co-segregating AFLPs as potential markers for marker- assisted selection	67
6 Abstract	70
7 Zusammenfassung	72
8 References	77

1 Introduction

1.1 Disease resistance in plants

1.1.1 Broad-spectrum resistance

Broad-spectrum resistance is a defence strategy that plants harbour against a large range of natural enemies such as bacteria, fungi, nematodes and viruses. Two types of broad-spectrum resistance, active and passive are distinguished. With respect to passive resistance, toxic compounds are constitutive independently of the presence of the pathogen (Osbourn et al., 1996). The toxic alkaloids in potato are an example of a passive resistance mechanism.

In case of active broad-spectrum resistance, defence reactions are initiated only when the plant is facing a pestilent attack; the effectiveness of such reactions is, nevertheless, against various potential pathogens. An example of active defence mechanism is the induced resistance (IR) that is activated e.g. upon an inducer-pathogen attack, enabling the plant to drive an effective defence response against a second attacking pathogen, the challenger. The first induced resistance is called local acquired resistance because it is limited to the infection site. The resistance may then spread systematically through the entire plant body leading to a broad-spectrum, systemic resistance (Ryals, 1996) such as the systemic acquired resistance (SAR). A set of genes known as SAR genes or pathogenesis related (PR) genes because of their implication in the production of the pathogenesis related proteins (PR-proteins) have been identified to be associated with SAR mechanisms in dicotyledonous species. However, it has not been clearly elucidated how the SAR is involved in monocotyledonous plants.

For plant improvement, broad-spectrum resistance has the advantage of being effective against several pathogen species. Breeding to increase the level of this resistance may be of great profit. However, the level of toxicity associated with the broad resistance mechanisms may have negative side effects particularly in food and fodder crops. For instance, it may be unacceptably high to humans and cattle, or decrease dramatically the nutritional value. Another disadvantage is that broad-spectrum resistance against a wide range of generalist pathogens and pests may be associated with increased attractiveness to specialist species (Niks and Lindhout, 2000).

1.1.2 Hypersensitivity resistance

The hypersensitive response (HR) is classically defined as a locally triggered cell death in the host plant at the site of attack by a pathogen (Agrios, 1997). Even hypersensitive resistance is, in many cases, associated with other active defence mechanisms, such as the PR-protein production, one particular aspect is that the HR effectiveness is race-specific. This enables the activation of the defence only to certain genotypes of the pathogen. For this reason, this type of resistance is also known as vertical resistance. In fact, a graph with many vertical columns is obtained when the reaction of resistance of a host plant genotype is plotted against a set of pathogen genotypes (VanderPlank, 1963).

To explain the race-specificity feature of hypersensitivity resistance, Flor (1971) proposed the hypothesis of a gene-for-gene interaction: 'For each gene conditioning resistance in the host, there is a specific gene conditioning pathogenicity in the parasite'. Flor made the emphasis on pathogenicity, literally virulence, but currently the emphasis is on avirulence. The resistance is determined by an interaction between the product of the resistance gene of the host plant and the product of the avirulence gene of the pathogen. A model elicitor/receptor has been suggested to elucidate the molecular basis of the gene-for-gene interaction (Keen, 1982).

In this model the dominant allele *R* for resistance produce a receptor molecule that recognizes an elicitor molecule produced by the dominant allele *Avr* of the avirulence gene. This recognition event elicits the hypersensitive reaction, which consists of a signal transduction pathway leading to a cascade of physiological reactions in the plant that are responsible for cell-death in the infection site (Blumwald et al., 1998). In case the pathogen carries the virulence allele, *avr*, rather than the avirulence allele, *Avr*, there will be no, or a mutilated, avirulence gene product and therefore, no recognition event will take place.

During the last years, the isolation of several *R* genes and the characterization of their products permitted considerable advances in the knowledge of the molecular basis of specific resistance. The comparison of different gene products revealed a strong sequence homology as well as five types of conserved structural domains: LRR (Leucine-Rich Repeats), Ser/Thr kinase (serine/threonine kinase), NBS (Nucleotide Binding Sites), LZ (Leucine Zipper) and TIR (Toll Interleukin Receptor). The *R* products seem to combine a receptor domain with an effector domain ensuring two main functions: the recognition of elicitor molecules thanks to protein-protein interaction